

Phenotypes in phylogeography: Species' traits, environmental variation, and vertebrate diversification

Kelly R. Zamudio^{a,1}, Rayna C. Bell^{b,c,d}, and Nicholas A. Mason^a

^aDepartment of Ecology and Evolutionary Biology, Cornell University, Ithaca, NY 14853; ^bMuseum of Vertebrate Zoology, University of California, Berkeley, CA 94720; ^cDepartment of Integrative Biology, University of California, Berkeley, CA 94720; and ^dDepartment of Vertebrate Zoology, National Museum of Natural History, Smithsonian Institution, Washington, DC 20560

Edited by Francisco J. Ayala, University of California, Irvine, CA, and approved May 18, 2016 (received for review February 10, 2016)

Almost 30 y ago, the field of intraspecific phylogeography laid the foundation for spatially explicit and genealogically informed studies of population divergence. With new methods and markers, the focus in phylogeography shifted to previously unrecognized geographic genetic variation, thus reducing the attention paid to phenotypic variation in those same diverging lineages. Although phenotypic differences among lineages once provided the main data for studies of evolutionary change, the mechanisms shaping phenotypic differentiation and their integration with intraspecific genetic structure have been underexplored in phylogeographic studies. However, phenotypes are targets of selection and play important roles in species performance, recognition, and diversification. Here, we focus on three questions. First, how can phenotypes elucidate mechanisms underlying concordant or idiosyncratic responses of vertebrate species evolving in shared landscapes? Second, what mechanisms underlie the concordance or discordance of phenotypic and phylogeographic differentiation? Third, how can phylogeography contribute to our understanding of functional phenotypic evolution? We demonstrate that the integration of phenotypic data extends the reach of phylogeography to explain the origin and maintenance of biodiversity. Finally, we stress the importance of natural history collections as sources of high-quality phenotypic data that span temporal and spatial axes.

phylogeography | phenotype | function | trait | concordance

Phylogeography, as originally defined, focused on processes governing the spatial distribution of genealogical lineages within species (1). One of the strengths of the field at its inception was formalizing conceptual links among heredity (processes at the level of individual pedigrees), divergence at the population level, and phylogenetic relationships among species (1). This analytical framework bridged microevolutionary processes acting within populations and macroevolutionary patterns at larger spatial and temporal scales. From the earliest applications, empirical phylogeographic studies described spatial patterns of genetic diversity and inferred underlying mechanisms, thus contributing to the explanatory and predictive power of the field (2). If most species show phylogeographic structure caused by landscape features that impede gene flow, then the geographic distribution of divergent lineages should coincide among species that coinhabit those landscapes. Further, phylogeographic breaks or contact zones should arise as lineages diverge allopatrically or come into secondary contact after divergence, respectively. This explicit prediction (1) resulted in a search for shared geographic patterns in genetic structure among species and the birth of comparative phylogeography (3, 4). Now, with thousands of taxon-specific phylogeographic studies published and synthesized in comparative studies (5, 6), we have learned a tremendous amount about the geography of genetic structure both within and among species.

Phylogeography has progressed rapidly in the last three decades, with new genetic markers (7, 8), analytical techniques (9), and synergies with landscape ecology and population genetics (10–12). Combined, these advances have revealed previously unrecognized genetic variation and its spatial and environmental correlates; however, phenotypic variation in those same diverging lineages

has not received the same attention. Phenotypic variation among populations across a species' range is common and often serves as the initial motivation for phylogeographic studies; however, most studies focus primarily on spatial variation in genetic lineages (7) and their distribution relative to environmental or geographic features of the landscape. Phenotypes are targets of selection and affect the performance of organisms in variable environments; combined, both processes contribute to diversification. Furthermore, different classes of phenotypes vary in how they impact processes such as dispersal, colonization, and persistence, thereby providing a window into the importance of various evolutionary processes in current and historical selective environments. Genetic structure of neutral genes, on the other hand, primarily reflects demographic processes (e.g., drift, expansion, changes in effective population size) that are a consequence of historical biotic and abiotic conditions during a species' evolutionary history. Thus, a new conceptual framework that explicitly integrates quantitative analyses of phenotypic variation within a phylogeographic framework can greatly enhance our knowledge of how genetic and phenotypic divergence arise, how they are linked, and how they respond to changing ecological and evolutionary contexts.

Here we review three research areas that exemplify the benefits of integrating phenotypic and genetic datasets in vertebrate phylogeography. First, we review how species-specific traits and their interactions with the environment predict concordant or idiosyncratic phylogeographic patterns among codistributed species. Second, we examine mechanisms that underlie the spatial concordance or discordance between phenotypic and genetic diversification. Third, we consider how phylogeography contributes to our understanding of functional phenotypic variation. For each topic, we describe case studies to highlight how the integration of phenotypic and genetic evolution has contributed to long-standing questions in evolutionary biology and has advanced our understanding of biodiversity. Finally, we emphasize the importance of natural history and field collections for the successful integration of organismal phenotypes and phylogeographic studies.

Species-Specific Traits and Idiosyncratic Phylogeographic Patterns

Comparative phylogeography seeks to characterize concordant phylogeographic breaks or contact zones, biogeographic “hot-spots” for understanding mechanisms shaping genetic structure within and among species with shared distributions (13, 14). A common assumption of comparative phylogeography is that taxa evolving in particular landscapes respond similarly to the abiotic and biotic elements that cause genetic divergence. We know,

This paper results from the Arthur M. Sackler Colloquium of the National Academy of Sciences, “In the Light of Evolution X: Comparative Phylogeography,” held January 8–9, 2016, at the Arnold and Mabel Beckman Center of the National Academies of Sciences and Engineering in Irvine, CA. The complete program and video recordings of most presentations are available on the NAS website at www.nasonline.org/ILE_X_Comparative_Phylogeography.

Author contributions: K.R.Z., R.C.B., and N.A.M. designed and wrote the paper.

The authors declare no conflict of interest.

This article is a PNAS Direct Submission.

¹To whom correspondence should be addressed. Email: kelly.zamudio@cornell.edu.

however, that species and populations vary in tolerance, plasticity, adaptive potential, and biotic interactions, all of which mediate responses to environmental variation (15–17) and ultimately dictate the degree of spatial and temporal concordance in genetic structure. The early definition of phylogeographic response categories acknowledged that differences could stem from species-specific traits such as dispersal potential and life history (1, 18). Not surprisingly, species that are exceptions to regional phylogeographic patterns have been identified in most, if not all, phylogeographic hotspots, precluding generalizations and challenging expectations for shared causes of organismal diversification.

Given that species-specific phenotypes can dictate spatial variation in population responses to environmental change, phylogeography would benefit from a more integrative and inclusive framework, one that incorporates predictions based on those phenotypes, an approach that has been termed “trait-based phylogeography” (19). A parallel example of trait-based approaches can be found in the emerging field of biodiversity and ecosystem function, which arose at the interface of community and ecosystem science (20). This new framework breaks from the view of species diversity as an epiphenomenon driven by a combination of abiotic environmental factors (e.g., temperature, rainfall, soil fertility), ecosystem processes that are themselves determined by these abiotic factors (e.g., productivity, biomass and nutrient cycling), and biotic interactions among species within communities (e.g., competition and predation). Instead, this new field considers biodiversity—in particular, the identity and diversity of species—as a driver of ecosystem functioning (20) and establishes causality between a species’ traits and the processes that in turn have functional consequences for ecosystems (20). Our current view of biodiversity in phylogeography parallels the “old view” in ecology by considering the genetic structure of species as a consequence of abiotic conditions and the evolutionary “function” of lineage births and deaths. In other words, species themselves, and their traits, are typically not considered as functionally causal in the processes that ultimately shape them. When we consider that traits can alter an organism’s demography and interactions with the environment, we can no longer ignore the dynamic nature of these interactions and their impact on lineage diversification (21). Thus, this paradigm shift challenges the expectation that temporal and spatial concordance among species should be the expected pattern in comparative phylogeography (22).

Phenotypes can either promote or constrain population divergence, depending on their function and interaction with the environment. For example, phenotypes that directly affect dispersal or persistence in new environments, such as those related to locomotor efficiency, physiological tolerance, or body size, can influence the frequency of migration and gene flow among subdivided populations. Others, such as recruitment rate, life span, and time to maturity will affect population size and turnover and thus the amount of genetic variation in subdivided populations. Finally, sexually selected phenotypes may not affect demography directly but can affect the distribution of genetic diversity indirectly via assortative mating, species recognition, and inbreeding avoidance. Variation in the distribution of phenotypes with different functions and the concordance of these phenotypes across species provide opportunities to quantify the importance of specific evolutionary processes for species inhabiting similar environments.

We are not implying that researchers have completely ignored species’ traits in interpreting phylogeographic patterns. Many studies consider dispersal capacity, environmental tolerance, and other characteristics that contribute to diversification (19, 23–26) or apply predictions derived from species-specific traits in a priori hypothesis testing (21, 27–29). To date, studies adopting a comparative trait-based framework typically have focused on groups of organisms evolving in and adapting to particular habitats. A comparison of four distantly related and allopatric temperate amphibian species demonstrated that population divergences are

significantly lower in two desert species that breed in ephemeral habitats than in two species inhabiting mesic forested landscapes (30). The stochastic persistence of breeding ponds across years in arid habitats may select against site fidelity and favor increased dispersal and larger physiological tolerances to inhospitable environments (30). A second study in the tropics confirmed that topographic complexity and especially macrohabitat preferences had strong effects on population divergence, so that species occupying forests and topographically complex regions showed deeper phylogeographic structure (31). Lower vagility across complex terrain and reliance on specific breeding habitats may lead to greater phylogeographic divergence in rainforest species. In contrast, species in more open landscapes typically use ephemeral and unpredictable breeding sites suitable for vagile generalists, possibly reducing intraspecific divergence (31).

Although trait-based analyses of shared phylogeographic structure yield important correlational evidence for divergence mechanisms (19, 26), the next step in this predictive framework is to examine species-specific traits that are selected for in particular landscapes and to quantify the extent to which those traits then contribute to diversification. An important advance in this direction is the development of model-based phylogeographic methods that incorporate phenotypic variation. These efforts stem from the realization that lack of concordance in temporal and spatial clustering in codistributed taxa may not mean that taxa are not responding to a common landscape or climatic barrier (32); rather, discrepancies may reflect variation in ecological traits and dispersal capabilities of taxa sampled across the presumed barrier (33). These efforts refine expectations for spatial concordance and temporally clustered divergences by explicitly including geography and trait-based responses for each species (27). A recent study examined trait-based phylogeographic predictions using flightless beetles that coinhabit the Cycladic Plateau in the central Aegean archipelago. The species differ in body size and associate with different soil types, both traits that affect dispersal capacity and persistence in habitat patches of different sizes (22). The authors investigated the relative importance of geographic factors and species-specific traits (soil-type preference, body size) on population divergence of 13 codistributed species. They found greater support for phylogeographic concordance when the null expectation of divergence times incorporated geographic and species-specific trait data (22). Efforts such as these to inform phylogeographic inferences with relevant differences among species have great potential for improving our understanding of how landscapes and species-specific traits interact during diversification (21, 34).

These empirical studies demonstrate that integrating species’ traits in phylogeographic studies can help explain the concordance (or discordance) of genealogical differentiation for species living in shared environments. This exciting prospect will greatly increase the impact of phylogeography in biodiversity science, and future studies need to consider explicitly how phenotypes can be incorporated into their predictive frameworks. Just as common responses to phylogeographic barriers became a null hypothesis early in the history of the field (33), an understanding of how organismal traits mediate responses to variable environments and demography will be necessary for a complete picture of the expected mode and rate of phylogeographic diversification (22). Achieving this understanding will require the development of metrics for quantifying phenotypic divergence and methods that explicitly incorporate those data in phylogeographic predictions and analyses (34). To be comparable, studies of geographic variation in phenotypes should strive to collect analogous data across species’ ranges. Doing so may be challenging, because phenotypes of interest will vary among systems, and each field has unique standards for replicable data, but these efforts will greatly expand the database of comparative traits for broader investigations of organismal divergence.

Evolutionary Mechanisms Linking Genetic and Phenotypic Diversification

All lineages harbor phenotypic and genetic variation among individuals, and this variation can be geographically partitioned in different ways (Fig. 1 and Table 1). In populations that remain geographically and genetically isolated, theory predicts that phenotypic differences will become more pronounced over time as the result of both neutral and selective processes (35). However, geographic variation in phenotypes does not always coincide with phylogeographic breaks (36). One prominent goal of phylogeography is to infer the biological mechanisms that partition genetic and phenotypic variation among populations. Recent advances in modeling (37), predictive frameworks for incorporating phenotypes into phylogeography (33), and DNA sequencing technologies (7) have bolstered our ability to identify and quantify the mechanisms that generate this diversity. Here, we illustrate those biological mechanisms that favor spatial concordance (or discordance) of genetic and phenotypic variation within a phylogeographic framework.

Geographically Clustered Phenotypes with Phylogeographic Structure. Phylogeographic structure often coincides with phenotypic variation (Fig. 1A) (35). In fact, many phylogeographic studies are initially motivated by phenotypic differences among lineages separated by putative barriers to gene flow. Phylogeographic structure is most pronounced when populations are separated by non-permeable barriers to gene flow and/or in taxa with limited capacity for dispersal across physical barriers (4). As a corollary,

those lineages with deeper phylogeographic structure likely have greater levels of phenotypic divergence, arising from longer periods of independent neutral and selective evolution (38). A recent study compared genetic and phenotypic differentiation between allopatric, ecologically similar sister species from eight genera of Peruvian passerine birds in cloud forests separated by the Marañón Valley, a prominent physical barrier to gene flow among central Andean taxa (39). Allospecies showed a positive association between genetic divergence and phenotypic differentiation, especially in plumage differentiation, a trait important for social signaling and species recognition. Empirical studies in treefrogs (40) and butterflyfishes (41) indicate that although the degree of phenotypic differentiation among vertebrates may scale with the strength and duration of genetic isolation, patterns can be highly idiosyncratic among taxa. Additionally, differences in the degree of phenotypic divergence among taxa or categories of traits can represent variation in evolutionary constraints or selective pressures driving the tempo and mode of phenotypic differentiation, which in turn can shape neutral genetic connectivity among populations.

Selective pressures vary temporally and geographically, thereby potentially altering the context-dependent fitness of phenotypic variants within lineages (42). If environmental conditions favor one phenotype, then populations may diverge phenotypically and genetically through local adaptation (43). The spatial arrangement of suitable habitat in heterogeneous landscapes, such as mosaics or clines, can also promote geographically clustered phenotypic variation (44). For example, strawberry poison dart frogs are highly polymorphic, and genetic distances among populations are

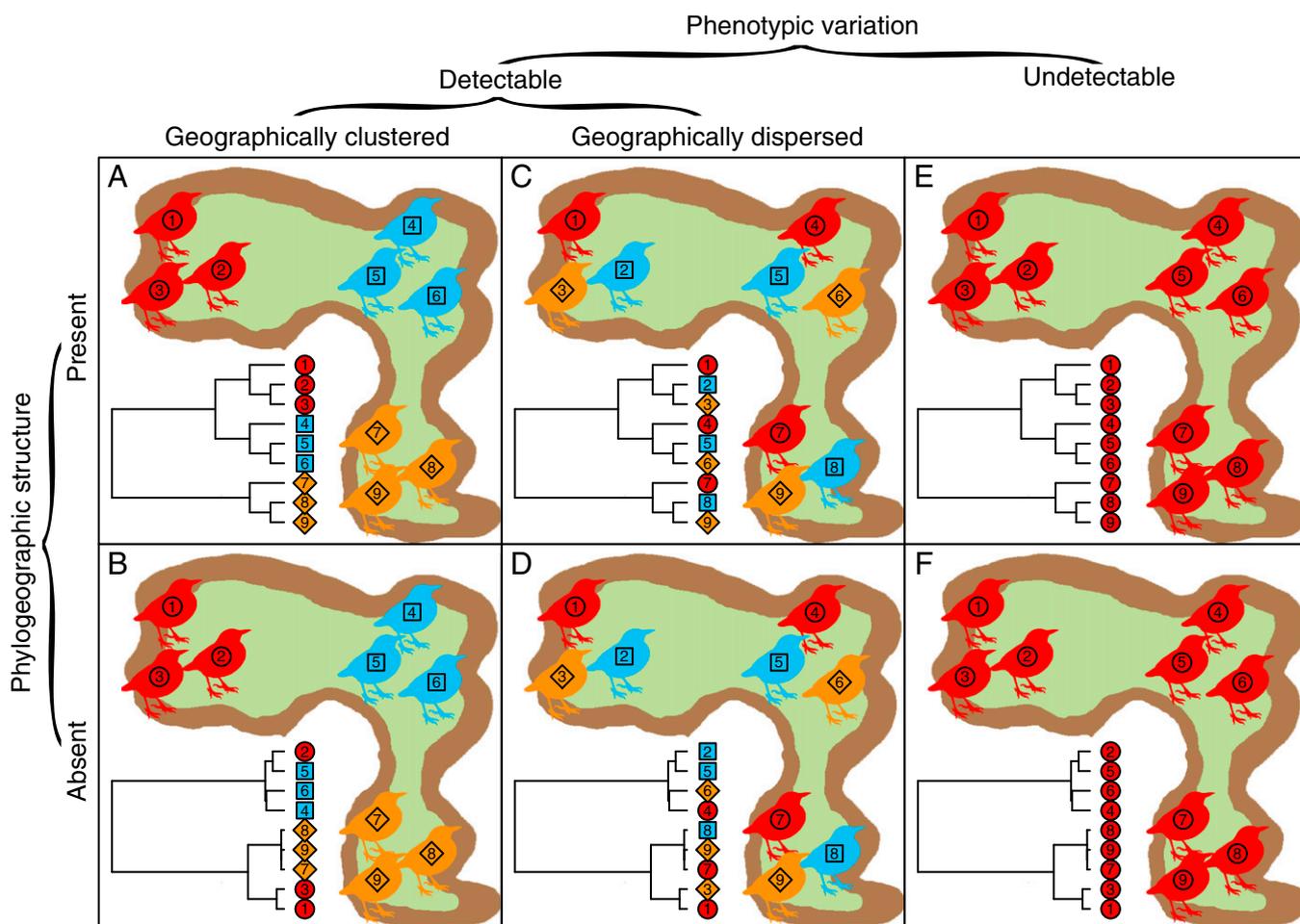


Fig. 1. Potential patterns of phylogeographic structure and phenotypic diversity among populations (A–F). Although discrete morphs are depicted here, similar patterns can arise for phenotypes with continuous variation.

Table 1. Patterns of phylogeographic structure and phenotypic diversity in vertebrates

Geographic variation in phenotypes	Phylogeographic structure	Biological mechanisms or processes	Empirical examples
Clustered	Present	Neutral divergence, local adaptation, divergent sexual selection	Neotropical oscines (39); treefrogs (40); butterflyfishes (41)
Clustered	Absent	Rapid and recent diversification, phenotypic plasticity, gene flow	Dark-eyed juncos (48); redpoll finches (52); perch and roach (53); spadefoot toads (54)
Dispersed	Present	Retention of ancestral polymorphism, convergent local adaptation, phenotypic plasticity, balancing selection	Side-blotched lizards (57); desert cichlids (58); threespine sticklebacks (61); trout (62); mollies (63); humans (64)
Dispersed	Absent	Convergent local adaptation, phenotypic plasticity, balancing selection, gene flow	Red crossbills (65, 66); ground snakes (67)
Undetectable	Present	Stabilizing selection, "cryptic" diversification	Sun skinks (70); black salamanders (71); plain-backed thrushes (72); field voles (73); rainforest skinks (75)
Undetectable	Absent	Stabilizing selection, gene flow	Straw-colored fruit bats (77)

For each of the patterns illustrated in Fig. 1 we include potential mechanisms contributing to the distribution of phenotypic and genetic variation and empirical examples.

more strongly associated with phenotypic differences than with geographic distances, suggesting a role for local adaptation related to predation and aposematism (25). Similarly, genetic isolation imposed by ecological variation also contributes to geographically clustered phenotypic and genealogical diversity in Mediterranean blue tits (45) and bats (46). Thus, a phylogeographic framework can reveal patterns of phenotypic and genetic variation and their regional concordance (or lack thereof), thereby elucidating roles of neutral and selective processes in lineage differentiation.

Geographically Clustered Phenotypes Without Phylogeographic Structure. In many systems, geographically clustered phenotypic variation exists without phylogeographic structure (Fig. 1*B*). Phenotypic differences can arise quickly through localized divergent selection, or, alternatively, they may seem to arise quickly because traditional molecular markers may not detect the phylogenetic structure underlying rapid diversification (47). Dark-eyed juncos, for example, exhibit pronounced phenotypic variation that is geographically clustered among subspecies that exhibit little to no phylogeographic structure (48). This pattern likely reflects subtle phylogeographic structure resulting from recent, rapid genetic isolation or adaptive divergence with ongoing gene flow among subspecies. Fortunately, high-throughput sequencing offers increased resolution and the capacity to distinguish between incomplete lineage sorting and ongoing gene flow, thereby improving our ability to infer ongoing biological processes in these cases (49).

Alternatively, clustered phenotypic variation among populations that lack phylogeographic structure may reflect biological processes, such as phenotypic plasticity (50, 51). Phenotypic plasticity involves developmental and phenotypic responses to different environmental regimes, thereby generating phenotypic diversity without genetic differentiation. For example, redpoll finches exhibit a gradient in which longer-billed individuals with streaking occur at lower latitudes, shorter-billed individuals with little to no streaking occur at higher latitudes, and many individuals express intermediate phenotypes (52). However, genome-scale SNP analyses revealed that redpolls constitute a single gene pool, regardless of their phenotype or geographic origin, so the paucity of genetic differentiation among phenotypes at a continental scale is probably not the result of recent divergence and/or insufficient molecular data (52). Furthermore, polygenic patterns of gene expression are strongly correlated with continuous variation in bill shape and plumage patterns, indicating possible roles for plasticity or variation in regulatory elements in generating and maintaining geographically clustered phenotypes (52). Recent studies in perch and roach (53) and spadefoot toads (54) similarly highlight the potential role of phenotypic plasticity in generating geographically clustered phenotypic variation without phylogeographic structure.

Distinguishing between recent, rapid bouts of adaptive genetic differentiation and phenotypic plasticity in natural populations can be difficult (55). Definitively demonstrating phenotypic plasticity requires experimental studies of captive populations or long-term longitudinal datasets, which are logistically challenging in many vertebrate, nonmodel species. Nonetheless, combining high-throughput sequencing with experimental studies to disentangle phenotypic plasticity and adaptive genetic divergence within a phylogeographic context is an exciting area of ongoing research.

Geographically Dispersed Phenotypes with Phylogeographic Structure. Many species are polymorphic with discrete or continuous phenotypic variation shared among phylogeographically structured populations (Fig. 1*C*). This pattern can be maintained through various evolutionary processes, including retention of ancestral polymorphism, balancing selection, parallel adaptation to locally variable conditions, and phenotypic plasticity. Multiple populations of side-blotched lizards in the western United States, for example, share polymorphism in throat color associated with different mating strategies that is maintained through negative frequency-dependent selection (56) and has persisted through multiple bouts of genetic isolation among populations (57).

Phenotypic plasticity also can contribute to geographically dispersed variation with phylogeographic structure. If plastic responses are retained among multiple populations that experience genetic isolation, then similar environmental conditions can result in shared polymorphism with phylogeographic structure. Desert cichlids restricted to the Cuatro Ciénegas valley in northern Mexico exhibit strong phylogeographic structure, and populations from each lagoon are genetically distinct from all others. However, both deep-bodied benthic morphs and slender-bodied limnetic morphs occur in each population, suggesting a role for phenotypic plasticity in generating and maintaining shared polymorphism among isolated populations (58).

Parallel adaptation, or the independent evolution of similar adaptive phenotypes in different populations, is another potential mechanism for shared phenotypic variation in species with marked phylogeographic structure. Parallel adaptations evolve under similar selective pressures, often acting on standing ancestral genetic variation (59), resulting in similar phenotypes among genetically isolated lineages (60). Prominent examples of parallel evolution include changes in the ectodysplasin signaling pathway in threespine sticklebacks that result in reduced armor plating in multiple, independent freshwater populations (61). Life history differentiation in trout (62), adaptations to sulphidic habitats in mollies (63), and lactase persistence in humans (64) underscore the potential for parallel evolution of shared phenotypic variation

among geographically and genetically isolated populations across a large diversity of taxa.

Geographically dispersed and shared phenotypic variation with phylogeographic structure (Fig. 1C) may arise through similar processes that generate geographically clustered phenotypes without phylogeographic structure (Fig. 1B), albeit at different temporal, spatial, and phylogenetic scales. Parallel adaptive and plastic responses to similar temporal or spatial variation in environmental conditions within the range of each lineage can generate shared polymorphisms among populations, even if they are separated by prolonged bouts of genetic isolation (Fig. 1C). Furthermore, rapid adaptation and phenotypic plasticity in response to regional conditions also can induce geographically clustered phenotypic variation without perceivable phylogeographic structure at larger spatial scales, especially if gene flow reduces genetic isolation among populations (Fig. 1B). Thus, the same adaptive and plastic processes can shape geographic and phylogenetic partitioning of phenotypic and genetic variation over space and time at different scales.

Geographically Dispersed Phenotypes Without Phylogeographic Structure. Phenotypic variation also can be dispersed and shared among populations that do not exhibit phylogeographic structure (Fig. 1D). Intuitively, this pattern can arise if processes that generate shared phenotypic variation (e.g., adaptations to spatial or temporal environmental heterogeneity, balancing selection, or phenotypic plasticity) occur among populations that readily exchange genes with one another. Polymorphism in red crossbills provides an empirical example: Their lower mandibles curve either left or right to facilitate feeding on conifer cones, and polymorphism is maintained by frequency-dependent selection in multiple populations connected by gene flow and thus weakly differentiated (65, 66). Likewise, highly polymorphic ground snakes include various color morphs that are present in multiple, genetically undifferentiated populations (67). Although discrete forms of polymorphism are perhaps easier to identify, variation in continuous phenotypes—such as body size or limb length—is also common among populations that lack phylogeographic structure. Dispersed polymorphism among populations that lack phylogeographic structure is more likely in vagile than dispersal-limited taxa, and the maintenance of phenotypic diversity will depend on the strength of selection acting on phenotypic variants among populations connected by high levels of gene flow.

Uniform Phenotypes with Phylogeographic Structure. Many lineages exhibit phylogeographic structure with little or no detectable phenotypic variation (Fig. 1E). In the case of strong stabilizing selection acting on traits that characterize a species' niche, populations will track suitable habitat as it appears and disappears over time (68). As such, ancestral populations can subdivide and accumulate genetic differences without morphological divergence, generating "cryptic" lineages or species (69). Philippine sun skinks, for example, exhibit deep phylogenetic splits concordant with geography but show little to no morphological variation among lineages (70). Similar patterns in black salamanders (71), plain-backed thrushes (72), and field voles (73) indicate that cryptic lineages are prevalent among vertebrates. Although cryptic lineages may not persist if previously isolated populations come into contact and exchange genes freely, speciation theory predicts that reproductive isolation—and postzygotic reproductive isolation in particular—increases with divergence time between lineages, in part through the accrual of Dobzhansky–Muller incompatibilities (74). Thus, study systems in which cryptic lineages occur in secondary contact provide an opportunity to address the evolution of postzygotic reproductive isolation between cryptic populations in nature. For example, divergence time and the degree of reproductive isolation are tightly correlated across five contact zones among cryptic lineages of rainforest skinks previously isolated in glacial refugia, indicating that, even in morphologically cryptic lineages, phylogeographic splits of increasing depth represent stages along the speciation continuum (75). Lineages with limited

dispersal and prominent evolutionary or developmental constraints are most likely to develop phylogeographic structure with no perceivable phenotypic diversity (69). We expect this pattern where strong biogeographic barriers to gene flow generate genetic divergence, and the selective environments between genetically isolated populations are relatively similar over space and time.

Uniform Phenotypes Without Phylogeographic Structure. Occasionally, vertebrate taxa do not vary perceivably in phenotypes within and among populations that do not also exhibit phylogeographic structure (Fig. 1F). Phenotypic uniformity among populations will be more likely if gene flow homogenizes populations (76). If environmental conditions are temporally and spatially consistent, then phenotypic variation may become fixed at an optimum, such that no phenotypic variation exists across a species' range (43). Furthermore, if individuals consistently disperse and exchange genes with other populations, phylogeographic structure will not accumulate over time. Straw-colored fruit bats in continental Africa, for example, display no phylogeographic structure and do not vary in phenotype across their expansive distribution (77). Vagile taxa that have expanded their range rapidly from a single glacial refugium, such as the blackpoll warbler (78), may display phenotypic uniformity and little to no phylogeographic structure among populations. A phylogeographic framework can disentangle cases in which continued gene flow homogenizes genetic and phenotypic variation among populations and cases in which historical demographic events, such as recent and/or rapid range expansions, have limited the time for phenotypic differences to accumulate (79).

As illustrated by the empirical and hypothetical examples discussed above (Fig. 1 and Table 1), the geographic partitioning of phenotypic and genotypic diversity is highly variable among vertebrates. Geographic partitioning of phenotypic variation across environmental gradients can occur with or without genetic differentiation, and in some cases the mechanisms for concordance (or lack thereof) are difficult to disentangle. Integrative approaches that combine high-throughput sequencing, experimental manipulations, and high-quality phenotypic datasets allow us to differentiate among biological mechanisms underlying phenotype–genotype concordance. Comparative studies adopting this framework will yield further examples of neutral divergence, local adaptation, and phenotypic and developmental plasticity, balancing selection and the prevalence of different evolutionary processes across taxa. Phylogeographic studies adopting this framework will also enhance our understanding of how rates and modes of phenotypic diversification vary among taxa. Finally, clarifying the evolutionary mechanisms underlying patterns of phenotypic and genetic diversity has implications for conserving biodiversity and for making accurate predictions of how species will respond to environmental change.

Phylogeography and Functional Variation in Phenotypes

Phylogeographic studies of functional variation in phenotypes have the potential to identify selective regimes that structure variation within and between species and ultimately shape the evolutionary history of functional traits. This approach can build on classic studies of hybrid zones and character evolution at macroevolutionary timescales by focusing on the spatial distribution of functional (or selected) traits within species. For instance, most hybrid zones result from secondary contact between populations or species that were previously allopatric; thus, the selective environment within the hybrid zone may not reflect the selective pressures that initiated divergence between parental lineages. Likewise, methods for quantifying character evolution above the species level typically do not account for within-species variation in phenotype and genotype (but see ref. 80) or the range of environmental conditions across a given species' distribution. Thus, a phylogeographic approach that encompasses phenotypic and environmental variation within species and contextualizes the demographic history of functional traits can provide exceptional insights into how organismal diversity evolves.

Identifying the role of extrinsic barriers in shaping the geographic distribution of functional phenotypic variation is an essential first step for investigating local adaptation. Thus, many studies quantify divergence at neutral genetic markers to investigate whether historical barriers coincide with the geographic distribution of variation in phenotypes such as coloration (25) or physiology (23). Within this historical context, phylogeographers then can investigate whether regional environmental variation and local adaptation contribute to phenotypic divergence (e.g., ref. 81) and identify instances of parallel phenotypic evolution among phylogeographic lineages that occupy similar selective environments in different geographic areas (82, 83). Even in study systems with only a single evolutionary origin of the trait of interest, a broader phylogeographic framework can inform the timing and/or direction of phenotypic change between genetically differentiated lineages. For example, conspicuous coloration and toxicity vary across the range of the granular poison-dart frog, and prevailing evolutionary theory contends that these traits should evolve in a correlated fashion. By reconstructing phylogeographic relationships across the species' range, Wang (84) demonstrated that the less conspicuous, more toxic population evolved from a more conspicuous, less toxic ancestor, thus challenging the view that conspicuousness and toxicity are tightly coupled. Finally, quantifying demographic processes, such as patterns of gene flow or changes in population size, and the distribution of phenotypic variation among divergent lineages can provide preliminary insights as to the strength of selection on adaptive phenotypes and whether gene flow between populations introduces adaptive phenotypes to new environments. For example, in the rock pocket mouse, in which melanic pelage evolves repeatedly on dark lava flows, high levels of gene flow between neighboring populations that differ phenotypically indicates that selection for color-matching is strong (82, 85). Furthermore, high rates of gene flow between melanic mice populations inhabiting neighboring lava flows suggests that on a finer spatial scale adaptive phenotypes in these different populations have a common genetic basis (82). Thus, characterizing geographic patterns of divergence at neutral genetic markers and in phenotypes contextualizes the demographic history of adaptive traits, and this evolutionary perspective then can inform in-depth investigations to identify the selective environment in which these traits evolved.

Phylogeographic studies of functional traits can identify the processes that shape adaptive variation and estimate the strength of selection acting on phenotypic variation by building links between locally adapted genotypes/phenotypes, population demography, and environmental variation in selective regimes. The challenge of this approach, however, is that it requires identifying and quantifying adaptive phenotypes, obtaining samples that encompass relevant environmental and phenotypic variation across the species' range, and characterizing functional genetic variation underlying adaptive phenotypes as well as neutral variation to estimate population demographic history. Given that the genetic architecture of adaptive phenotypes is unknown in most nonmodel vertebrate taxa, this approach has been applied primarily in systems with extensive genomic resources (e.g., the threespine stickleback; 86) or in systems in which the links between a particular adaptive phenotype and the underlying functional genetic variation are well defined. These systems include adaptive shifts in coloration caused by variation in genes encoding the melanin pathway (85), physiological adaptation to high-altitude environments caused by variation in hemoglobin subunit genes (87), tetrodotoxin resistance caused by variation in skeletal muscle sodium channels (88), and differences in adaptive immunity caused by variations in genes encoding the MHC class II subunits (89, 90).

One common result of functional-trait studies is the identification of recurrent novel mutations underlying similar phenotypes (88, 90, 91). For example, mutations in the melanocortin-1 receptor gene (*Mcr1*) are highly correlated with adaptive melanism in an Arizona population of rock pocket mice, but melanic populations in New Mexico show no association with variation at

Mcr1, indicating that their dark coloration must result from changes at different genes (82, 91, 92). This finding is in contrast to traits associated with threespine sticklebacks in which the same adaptive alleles underlie multiple independent freshwater invasions on a regional scale (86). A second theme is the role of gene flow in promoting adaptive evolution in some contexts (82) and impeding adaptation in others (89, 90). Finally, geographic sampling that captures environmental variation is especially valuable for understanding complex adaptive scenarios such as host-pathogen dynamics. Across its range, the lowland leopard frog exhibits population-level variation in survival after infection by the fungal pathogen *Batrachochytrium dendrobatidis*, which causes chytridiomycosis, a disease implicated in population declines or extinction in hundreds of amphibian species worldwide (93). Variation in immunity loci determines susceptibility to the pathogen such that several MHC alleles are strongly associated with increased survival or susceptibility in both experimental (94) and natural settings (90). These associations are decoupled, however, for populations surrounding a thermal spring, where warm water shields frogs from developing high pathogen loads but also precludes selection from increasing the frequency of MHC survival alleles (90).

In summary, quantifying functional genetic variation within the context of the phylogeographic history of a species and across the range of environments it inhabits can reveal how regional variation in selective regimes and demographic processes drives the evolution of adaptive phenotypes. Just as phylogeography initially formalized conceptual links among heredity, population divergence, and phylogenetic relationships among species, an analytical framework that advocates genealogical and spatially explicit analyses of intraspecific functional genetic and phenotypic variation will bridge microevolutionary processes acting on individual populations and macroevolutionary patterns at larger spatial and temporal scales. These integrative and rigorous approaches have been possible only in select systems to date, but identifying the underlying genetic basis of phenotypic variation within species is becoming increasingly tractable in vertebrates. Clearly, examining functional phenotypic variation in a phylogeographic framework holds great promise for exploring links between genotypic and phenotypic diversity and adaptation across variable environments.

Field Studies and Natural History Collections: Sources of Phenotypic Data

Organismal phenotypes, many of which are the target of selection and play important roles in species performance in variable environments, are important components of how we identify and categorize biodiversity. We have argued that, despite the clear benefits of integrating patterns of phenotypic evolution into phylogeographic predictions and inferences, this integration has yet to be fully realized. One reason is that high-quality phenotypic data are difficult to obtain; establishing a phenotypic database with robust sample sizes and fine-scale spatial sampling can be laborious. This challenge is solvable by relying on well-established methods in biodiversity science.

For centuries, naturalists relied solely on phenotypes to document diversity, study the relationship of organisms with their environment, and infer evolutionary change. Unaware of the genetic underpinnings of those phenotypes, early naturalists focused on explaining phenotypic diversity not only among but also within species. They did so with a large number of phenotypes, including behavior, color, morphology, life history, and ecological traits, among others. Fortunately, many of those efforts are archived in publications or are preserved in natural history collections, providing a sample of Earth's biota that typically extends back to the 19th century, and often includes representative coverage of species' distributions (95, 96). For some kinds of phenotypes, such as certain aspects of morphology preserved in museum specimens, natural history collections are a rich source of phenotypic data. In contrast, for phenotypes that are not easily preserved—such as behavior, ecological associations, or

physiological parameters—field studies will be the ultimate source, although the metadata associated with many preserved specimens often contain important information on behavior, habitat preferences, and other ecological associations (95).

Natural history collections also will have a large role in the analysis of more recent evolutionary change in phenotypes (95). For some species, long-term series of collections offer a unique opportunity to infer ancestral phenotypes and how those have changed with documented changes in the environment (97). Examining these data within a phylogeographic framework provides the evolutionary context to identify range-wide dynamics of phenotypic change and may highlight regional sources of adaptive variation. Furthermore, many of these phenotypic changes can be tied to strong selection imposed by changing environments, thus providing the link between genetic and phenotypic changes under different environmental contexts. These phenotype-genotype associations provide a mechanistic basis for inferring past changes at both recent and longer evolutionary time frames and a predictive framework for understanding how organisms will adapt to future natural and anthropogenic global change. Phenotypes, genetic structure, and environmental characteristics are intimately coupled in the processes of organismal divergence; thus advances in all three fields will enable the integrative study of divergences in natural populations.

Conclusions

We see great promise in ongoing methodological and conceptual advances that explicitly incorporate trait evolution in phylogeographic predictions and inferences. Our goal here was to highlight the

many valuable avenues for future work in this area. The field of phylogeography has changed since its origin, incorporating new techniques, new analyses, and increasingly, different sources of data. Although only a fraction of extant taxa have been surveyed, the field has revealed many common patterns and mechanisms underlying diversification within broadly divergent taxonomic groups. Meanwhile, our ability to quantify genetic and phenotypic variation also has expanded; thus, the field of phylogeography now is poised for another integration, this time by incorporating data on phenotypic variation in diverging lineages, understanding the selective and genetic basis for that variation, and quantifying the role that phenotypes play in diversification. This integration has the potential to unify once again disparate fields in evolutionary biology, and address how interactions among abiotic landscapes features and biological features of species shape biodiversity (98). This integrative framework is a powerful tool for understanding the effects of past global change on current biodiversity and for predicting the adaptive potential and resilience of species adapting to novel environments of the future.

ACKNOWLEDGMENTS. We thank J. Avise and F. Ayala for the invitation to participate in the Sackler Symposium *In the Light of Evolution X: Comparative Phylogeography*; and the K.R.Z., Lovette, Searle, and McGuire laboratories, A. Corl, A. Chavez, H. Greene, and two anonymous reviewers for constructive feedback on the manuscript. Our work is funded by National Science Foundation Research Grants DEB-0542848, DEB-1601072, and DEB-1309171; EPA Science to Achieve Results Fellowship F13F21201 (to N.A.M.); and a University of California Chancellor's Postdoctoral Fellowship (to R.C.B.).

- Avise JC, et al. (1987) Intraspecific phylogeography: The mitochondrial DNA bridge between population genetics and systematics. *Annu Rev Ecol Syst* 18:489–522.
- Buckley D (2009) Toward an organismal, integrative, and iterative phylogeography. *BioEssays* 31(7):784–793.
- Schneider CJ, Cunningham M, Moritz C (1998) Comparative phylogeography and the history of endemic vertebrates in the Wet Tropics rainforests of Australia. *Mol Ecol* 7(4):487–498.
- Hewitt G (2000) The genetic legacy of the Quaternary ice ages. *Nature* 405(6789):907–913.
- Beheregaray LB (2008) Twenty years of phylogeography: The state of the field and the challenges for the Southern Hemisphere. *Mol Ecol* 17(17):3754–3774.
- Camargo A, Sinervo B, Sites JW, Jr (2010) Lizards as model organisms for linking phylogeographic and speciation studies. *Mol Ecol* 19(16):3250–3270.
- Edwards SV, Shultz AJ, Campbell-Staton C (2015) Next-generation sequencing and the expanding domain of phylogeography. *Folia Zool (Brno)* 64(3):187–206.
- Garrick RC, et al. (2015) The evolution of phylogeographic data sets. *Mol Ecol* 24(6):1164–1171.
- Hickerson MJ, et al. (2010) Phylogeography's past, present, and future: 10 years after Avise, 2000. *Mol Phylogenet Evol* 54(1):291–301.
- Chan LM, Brown JL, Yoder AD (2011) Integrating statistical genetic and geospatial methods brings new power to phylogeography. *Mol Phylogenet Evol* 59(2):523–537.
- Hickerson MJ, Stahl EA, Lessios HA (2006) Test for simultaneous divergence using approximate Bayesian computation. *Evolution* 60(12):2435–2453.
- Oaks JR (2014) An improved approximate-Bayesian model-choice method for estimating shared evolutionary history. *BMC Evol Biol* 14:150.
- Rissler LJ, Smith WH (2010) Mapping amphibian contact zones and phylogeographical break hotspots across the United States. *Mol Ecol* 19(24):5404–5416.
- Moritz C, et al. (2009) Identification and dynamics of a cryptic suture zone in tropical rainforest. *Proc Biol Sci* 276(1660):1235–1244.
- Bernardo J, Spotila JR (2006) Physiological constraints on organismal response to global warming: Mechanistic insights from clinally varying populations and implications for assessing endangerment. *Biol Lett* 2(1):135–139.
- Satler JD, Zellmer AJ, Carstens BC (2016) Biogeographic barriers drive co-diversification within associated eukaryotes of the *Sarracenia alata* pitcher plant system. *PeerJ* 4:e1576.
- Ridenhour BJ, Brodie ED, Jr, Brodie ED, III (2007) Patterns of genetic differentiation in *Thamnophis* and *Taricha* from the Pacific Northwest. *J Biogeogr* 34(4):724–735.
- Avise JC, Helfman GS, Saunders NC, Hales LS (1986) Mitochondrial DNA differentiation in North Atlantic eels: Population genetic consequences of an unusual life history pattern. *Proc Natl Acad Sci USA* 83(12):4350–4354.
- Paz A, Ibáñez R, Lips KR, Crawford AJ (2015) Testing the role of ecology and life history in structuring genetic variation across a landscape: A trait-based phylogeographic approach. *Mol Ecol* 24(14):3723–3737.
- Loreau M (2010) Linking biodiversity and ecosystems: Towards a unifying ecological theory. *Philos Trans R Soc Lond B Biol Sci* 365(1537):49–60.
- He Q, Edwards DL, Knowles LL (2013) Integrative testing of how environments from the past to the present shape genetic structure across landscapes. *Evolution* 67(12):3386–3402.
- Papadopoulou A, Knowles LL (2015) Species-specific responses to island connectivity cycles: Refined models for testing phylogeographic concordance across a Mediterranean Pleistocene Aggregate Island Complex. *Mol Ecol* 24(16):4252–4268.
- Moritz C, et al. (2012) Integrating phylogeography and physiology reveals divergence of thermal traits between central and peripheral lineages of tropical rainforest lizards. *Philos Trans R Soc Lond B Biol Sci* 367(1596):1680–1687.
- Bell RC, et al. (2012) Comparative multi-locus phylogeography confirms multiple vicariance events in co-distributed rainforest frogs. *Proc Biol Sci* 279(1730):991–999.
- Wang JJ, Summers K (2010) Genetic structure is correlated with phenotypic divergence rather than geographic isolation in the highly polymorphic strawberry poison-dart frog. *Mol Ecol* 19(3):447–458.
- Smith BT, et al. (2014) The drivers of tropical speciation. *Nature* 515(7527):406–409.
- Massatti R, Knowles LL (2014) Microhabitat differences impact phylogeographic concordance of codistributed species: Genomic evidence in montane sedges (*Carex* L.) from the Rocky Mountains. *Evolution* 68(10):2833–2846.
- Brown JL, Knowles LL (2012) Spatially explicit models of dynamic histories: Examination of the genetic consequences of Pleistocene glaciation and recent climate change on the American Pika. *Mol Ecol* 21(15):3757–3775.
- Brumfield RT, Capparella AP (1996) Historical diversification of birds in northwestern South America: A molecular perspective on the role of vicariant events. *Evolution* 50(4):1607–1624.
- Chan LM, Zamudio KR (2009) Population differentiation of temperate amphibians in unpredictable environments. *Mol Ecol* 18(15):3185–3200.
- Rodríguez A, et al. (2015) Genetic divergence in tropical anurans: Deeper phylogeographic structure in forest specialists and in topographically complex regions. *Evol Ecol* 29(5):765–785.
- Oaks JR, et al. (2013) Evidence for climate-driven diversification? A caution for interpreting ABC inferences of simultaneous historical events. *Evolution* 67(4):991–1010.
- Papadopoulou A, Knowles LL (2016) Toward a paradigm shift in comparative phylogeography driven by trait-based hypotheses. *Proc Natl Acad Sci USA*, 10.1073/pnas.1601069113.
- Knowles LL, Alvarado-Serrano DF (2010) Exploring the population genetic consequences of the colonization process with spatio-temporally explicit models: Insights from coupled ecological, demographic and genetic models in montane grasshoppers. *Mol Ecol* 19(17):3727–3745.
- Gould SJ, Johnston RF (1972) Geographic variation. *Annu Rev Ecol Syst* 3:457–498.
- Endler JA (1973) Gene flow and population differentiation. *Science* 179(4070):243–250.
- Thomé MTC, Carstens BC (2016) Phylogeographic model selection leads to insight into the evolutionary history of four-eyed frogs. *Proc Natl Acad Sci USA*, 10.1073/pnas.1601064113.
- Lande R (1980) Genetic variation and phenotypic evolution during allopatric speciation. *Am Nat* 116(4):463–479.
- Winger BM, Bates JM (2015) The tempo of trait divergence in geographic isolation: Avian speciation across the Marañón Valley of Peru. *Evolution* 69(3):772–787.
- Warwick AR, Travis J, Lemmon EM (2015) Geographic variation in the Pine Barrens Treefrog (*Hyla andersonii*): Concordance of genetic, morphometric and acoustic signal data. *Mol Ecol* 24(13):3281–3298.

41. Waldrop E, et al. (2016) Phylogeography, population structure and evolution of coral-eating butterflyfishes (Family Chaetodontidae, genus *Chaetodon*, subgenus *Corallochaetodon*). *J Biogeogr* 43(6):1116–1129.
42. Wang JJ, Bradburd GS (2014) Isolation by environment. *Mol Ecol* 23(23):5649–5662.
43. Kawecki TJ, Ebert D (2004) Conceptual issues in local adaptation. *Ecol Lett* 7(12):1225–1241.
44. Forester BR, Jones MR, Joost S, Landguth EL, Lasky JR (2016) Detecting spatial genetic signatures of local adaptation in heterogeneous landscapes. *Mol Ecol* 25(1):104–120.
45. Charmantier A, Doutrelant C, Dubuc-Messier G, Fargevielle A, Szulkin M (2015) Mediterranean blue tits as a case study of local adaptation. *Evol Appl* 9(1):135–152.
46. Morales A, Villalobos F, Velazco PM, Simmons NB, Piñero D (2016) Environmental niche drives genetic and morphometric structure in a widespread bat. *J Biogeogr* 43(5):1057–1068.
47. Edwards S, Bensch S (2009) Looking forwards or looking backwards in avian phylogeography? A comment on Zink and Barrowclough 2008. *Mol Ecol* 18(14):2930–2933, discussion 2934–2936.
48. McCormack JE, et al. (2012) Next-generation sequencing reveals phylogeographic structure and a species tree for recent bird divergences. *Mol Phylogenet Evol* 62(1):397–406.
49. Edwards SV, Potter S, Schmitt CJ, Bragg JG, Moritz C (2016) Reticulation, divergence, and the phylogeography–phylogenetics continuum. *Proc Natl Acad Sci USA*, 10.1073/pnas.1601066113.
50. Crispo E (2008) Modifying effects of phenotypic plasticity on interactions among natural selection, adaptation and gene flow. *J Evol Biol* 21(6):1460–1469.
51. Pfennig DW, et al. (2010) Phenotypic plasticity's impacts on diversification and speciation. *Trends Ecol Evol* 25(8):459–467.
52. Mason NA, Taylor SA (2015) Differentially expressed genes match bill morphology and plumage despite largely undifferentiated genomes in a Holarctic songbird. *Mol Ecol* 24(12):3009–3025.
53. Faulks L, Svanbäck R, Eklöv P, Östman Ö (2015) Genetic and morphological divergence along the littoral–pelagic axis in two common and sympatric fishes: Perch, *Perca fluviatilis* (Percidae) and roach, *Rutilus rutilus* (Cyprinidae). *Biol J Linn Soc Lond* 114(4):929–940.
54. Rice AM, Pfennig DW (2010) Does character displacement initiate speciation? Evidence of reduced gene flow between populations experiencing divergent selection. *J Evol Biol* 23(4):854–865.
55. Merilä J, Hendry AP (2014) Climate change, adaptation, and phenotypic plasticity: The problem and the evidence. *Evol Appl* 7(1):1–14.
56. Sinervo B, Svensson E, Comendant T (2000) Density cycles and an offspring quantity and quality game driven by natural selection. *Nature* 406(6799):985–988.
57. Corl A, Davis AR, Kuchta SR, Sinervo B (2010) Selective loss of polymorphic mating types is associated with rapid phenotypic evolution during morphic speciation. *Proc Natl Acad Sci USA* 107(9):4254–4259.
58. Magalhaes IS, Ornelas-Garcia CP, Leal-Cardin M, Ramirez T, Barluenga M (2015) Untangling the evolutionary history of a highly polymorphic species: Introgressive hybridization and high genetic structure in the desert cichlid fish *Herichthys minckleyi*. *Mol Ecol* 24(17):4505–4520.
59. Barrett RD, Schluter D (2008) Adaptation from standing genetic variation. *Trends Ecol Evol* 23(1):38–44.
60. Stern DL (2013) The genetic causes of convergent evolution. *Nat Rev Genet* 14(11):751–764.
61. Colosimo PF, et al. (2005) Widespread parallel evolution in sticklebacks by repeated fixation of Ectodysplasin alleles. *Science* 307(5717):1928–1933.
62. Pearse DE, Miller MR, Abadia-Cardoso A, Garza JC (2014) Rapid parallel evolution of standing variation in a single, complex, genomic region is associated with life history in steelhead/rainbow trout. *Proc Biol Sci* 281(1783):20140012.
63. Pfenniger M, et al. (2015) Unique evolutionary trajectories in repeated adaptation to hydrogen sulphide-toxic habitats of a neotropical fish (*Poecilia mexicana*). *Mol Ecol* 24(21):5446–5459.
64. Tishkoff SA, et al. (2007) Convergent adaptation of human lactase persistence in Africa and Europe. *Nat Genet* 39(1):31–40.
65. Benkman CW (1996) Are the ratios of bill crossing morphs in crossbills the result of frequency-dependent selection? *Evol Ecol* 10(1):119–126.
66. Parchman TL, Benkman CW, Britch SC (2006) Patterns of genetic variation in the adaptive radiation of New World crossbills (Aves: *Loxia*). *Mol Ecol* 15(7):1873–1887.
67. Cox CL, Davis Rabosky AR (2013) Spatial and temporal drivers of phenotypic diversity in polymorphic snakes. *Am Nat* 182(2):E40–E57.
68. Wiens JJ, et al. (2010) Niche conservatism as an emerging principle in ecology and conservation biology. *Ecol Lett* 13(10):1310–1324.
69. Bickford D, et al. (2007) Cryptic species as a window on diversity and conservation. *Trends Ecol Evol* 22(3):148–155.
70. Barley AJ, Monahan PJ, Thomson RC, Grismer LL, Brown RM (2015) Sun skink landscape genomics: Assessing the roles of micro-evolutionary processes in shaping genetic and phenotypic diversity across a heterogeneous and fragmented landscape. *Mol Ecol* 24(8):1696–1712.
71. Reilly SB, Wake DB (2015) Cryptic diversity and biogeographical patterns within the black salamander (*Aneides flavipunctatus*) complex. *J Biogeogr* 42(2):280–291.
72. Alström P, et al. (2016) Integrative taxonomy of the Plain-backed Thrush (*Zoothera mollissima*) complex (Aves, Turdidae) reveals cryptic species, including a new species. *Avian Research* 7(1):1–39.
73. Paupério J, et al. (2012) Cryptic speciation in the field vole: A multilocus approach confirms three highly divergent lineages in Eurasia. *Mol Ecol* 21(24):6015–6032.
74. Bolnick DI, Near TJ (2005) Tempo of hybrid inviability in centrarchid fishes (Teleostei: Centrarchidae). *Evolution* 59(8):1754–1767.
75. Singhal S, Moritz C (2013) Reproductive isolation between phylogeographic lineages scales with divergence. *Proc Biol Sci* 280(1772):20132246.
76. Lenormand T (2002) Gene flow and the limits to natural selection. *Trends Ecol Evol* 17(4):183–189.
77. Peel AJ, et al. (2013) Continent-wide panmixia of an African fruit bat facilitates transmission of potentially zoonotic viruses. *Nat Commun* 4:2770.
78. Ralston J, Kirchman JJ (2012) Continent-scale genetic structure in a boreal forest migrant, the Blackpoll Warbler (*Setophaga striata*). *Auk* 129(3):467–478.
79. Marko PB, Hart MW (2011) The complex analytical landscape of gene flow inference. *Trends Ecol Evol* 26(9):448–456.
80. Revell LJ, Graham Reynolds R (2012) A new Bayesian method for fitting evolutionary models to comparative data with intraspecific variation. *Evolution* 66(9):2697–2707.
81. Ng J, Landeen EL, Logsdon RM, Gior RE (2013) Correlation between *Anolis* lizard dewlap phenotype and environmental variation indicates adaptive divergence of a signal important to sexual selection and species recognition. *Evolution* 67(2):573–582.
82. Hoekstra HE, Krenz JG, Nachman MW (2005) Local adaptation in the rock pocket mouse (*Chaetodipus intermedius*): Natural selection and phylogenetic history of populations. *Heredity (Edinb)* 94(2):217–228.
83. Richmond JQ, Reeder TW (2002) Evidence for parallel ecological speciation in scincid lizards of the *Eumeces skiltonianus* species group (Squamata: Scincidae). *Evolution* 56(7):1498–1513.
84. Wang JJ (2011) Inversely related aposematic traits: Reduced conspicuousness evolves with increased toxicity in a polymorphic poison-dart frog. *Evolution* 65(6):1637–1649.
85. Hoekstra HE, Drumm KE, Nachman MW (2004) Ecological genetics of adaptive color polymorphism in pocket mice: Geographic variation in selected and neutral genes. *Evolution* 58(6):1329–1341.
86. Deagle BE, Jones FC, Absher DM, Kingsley DM, Reimchen TE (2013) Phylogeography and adaptation genetics of stickleback from the Haida Gwaii archipelago revealed using genome-wide single nucleotide polymorphism genotyping. *Mol Ecol* 22(7):1917–1932.
87. Bulgarella M, et al. (2012) Multilocus coalescent analysis of haemoglobin differentiation between low- and high-altitude populations of crested ducks (*Lophonetta specularioides*). *Mol Ecol* 21(2):350–368.
88. Feldman CR, Brodie ED, Jr, Brodie ED, 3rd, Pfrender ME (2009) The evolutionary origins of beneficial alleles during the repeated adaptation of garter snakes to deadly prey. *Proc Natl Acad Sci USA* 106(32):13415–13420.
89. Savage AE, Becker CG, Zamudio KR (2015) Linking genetic and environmental factors in amphibian disease risk. *Evol Appl* 8(6):560–572.
90. Savage AE, Zamudio KR (2016) Adaptive tolerance to a pathogenic fungus drives major histocompatibility complex evolution in natural amphibian populations. *Proc Biol Sci* 283(1827):20153115.
91. Nachman MW, Hoekstra HE, D'Agostino SL (2003) The genetic basis of adaptive melanism in pocket mice. *Proc Natl Acad Sci USA* 100(9):5268–5273.
92. Hoekstra HE, Nachman MW (2003) Different genes underlie adaptive melanism in different populations of rock pocket mice. *Mol Ecol* 12(5):1185–1194.
93. Fisher MC, et al. (2012) Emerging fungal threats to animal, plant and ecosystem health. *Nature* 484(7393):186–194.
94. Savage AE, Zamudio KR (2011) MHC genotypes associate with resistance to a frog-killing fungus. *Proc Natl Acad Sci USA* 108(40):16705–16710.
95. Holmes MW, et al. (2016) Natural history collections as windows on evolutionary processes. *Mol Ecol* 25(4):864–881.
96. Graham CH, Ferrier S, Huettman F, Moritz C, Peterson AT (2004) New developments in museum-based informatics and applications in biodiversity analysis. *Trends Ecol Evol* 19(9):497–503.
97. Özgo M, Schilthuizen M (2012) Evolutionary change in *Cepaea nemoralis* shell colour over 43 years. *Glob Change Biol* 18(1):74–81.
98. Greene HW (2005) Organisms in nature as a central focus for biology. *Trends Ecol Evol* 20(1):23–27.